



Protein A/G Immunoassay (co) Precipitation Kit (Gel method)
PRODUCT DATA SHEET

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Cat No: AKIT-Protein A/G

Description

The Protein A/G immunoprecipitation kit (gel method) contains enough reagents to complete 40 reactions using 25 μ L gel for each reaction. It can effectively complete antigen immunoprecipitation (IP) and immunocoprecipitation (Co-IP) experiments, and the antibody dosage is less than 10 μ g.

Protein A/G agarose gel provided in the kit can realize rapid and convenient antigen separation. The immunoprecipitation kit is equipped with optimized prefabricated buffer, which provides the best reaction conditions for the immunoprecipitation experiment and enhances the stability of the immunoprecipitation experiment. This product can be widely used in the immunoprecipitation reaction of antigens in cell lysate, cell secreted supernatant, serum, ascites and other samples.

For custom sizes, formulations or bulk quantities please contact our customer service department.

Website: www.abvigen.com **Phone:** +1 929-202-3014 **Email:** info@abvigenus.com

Kit Composition

Protein A/G agarose gel	1 mL
IP Lysis/Wash Buffer	100 mL
SDS-PAGE Sample Loading Buffer (5 \times)	10 mL
Phosphatase inhibitor cocktail (50 \times)	2 mL
PMSF (100 \times)	1 mL
Elution Buffer	5 mL
Neutralization Buffer	1 mL



Operation Process

Adherent cell sample:

1. Remove the medium and wash the cells twice with PBS.
2. Cells were collected into 1.5 mL EP tube and IP Lysis/Wash Buffer was added proportioned, and add corresponding inhibitor such as PMSF. After mixing, the cells were placed on ice for 5 ~ 20 min (mixed several times).
3. At 4°C, 12000 ~ 16000 xg, centrifuge for 10 min to collect the supernatant and place it on ice for subsequent experiments (or place it at -80°C for long-term storage).

Petri dish IP Lysis/Wash Buffer Recommended volume:

Petri dish size/surface area	Volume of IP Lysis/Wash Buffer
100 mm x 100 mm	500 ~ 1000 μ L
100 mm x 60 mm	100 ~ 300 μ L
6-well plate	100 ~ 200 μ L

Suspended cell sample:

1. Cells were collected at 4°C, 500 ~ 1000 xg, 10 min, and the supernatant was discarded.
2. Wash the cells with PBS once, that is, suspend the cell mass with PBS at 4°C, 500 ~ 1000 xg, 5 min, collect the cells, discard the supernatant.
3. Resuspend cells with pre-cooled IP Lysis/Wash Buffer. The 500 μ L IP Lysis/Wash Buffer was used for every 50 mg cells. At the same time, add the corresponding inhibitors such as PMSF, mix well and put on the ice for 5 ~ 20 min (mix several times during the period).
4. At 4°C, 12000 ~ 16000 xg, centrifuge for 10 min to collect the supernatant and place it on ice for subsequent experiments (or place it at -80°C for long-term storage).

Serum sample:

It is recommended to dilute a serum sample with IP Lysis/Wash Buffer to a final concentration of target protein of 50 to 150 μ g/mL and place it on ice for future use (or place it at -20°C for long-term storage).

Preparation of immune complex

Note: The amount and incubation time required for the sample are dependent on each specific antibody antigen system and may need to be optimized for maximum yield.

The following protocol is for 2 ~ 10 μ g affinity purified antibodies, which can be scaled up as required.



1. In a centrifuge tube, the cell lysate of each sample was combined with 2 ~ 10 μ g immunoprecipitation antibody. The recommended total protein amount for each immunoprecipitation reaction is 500 ~ 1500 μ g.
2. Dilute the antibody and prepared sample to 300 ~ 500 μ L from IP Lysis/Wash Buffer.
3. Incubate at room temperature for 1 ~ 2 h, or 4°C for 2 ~ 4 h to form immune complexes.

Immunoprecipitation:

Note: In order to ensure the uniform distribution of gel, mix the gel in the bottle by repeatedly reversing or slightly vortexing before use.

1. Add 25 μ L of Protein A/G agarose gel into 1.5 mL centrifuge tube.
2. Add 500 μ L pre-cooled PBS to the gel and mix gently.
3. Put the centrifuge tube into the centrifuge at 1000 rpm for 5 min to collect the gel to the bottom of the centrifuge tube and remove the supernatant.
4. Add 200 ~ 500 μ L IP Lysis/Wash Buffer to the centrifuge tube. Reverse the centrifugal tube several times or mix with a slight vortex for 1 min. Put the centrifuge tube into the centrifuge at 1000 rpm for 5 min to collect the gel to the bottom of the centrifuge tube and remove the supernatant.
5. The antigen sample/antibody mixture was added to the centrifuge tube with gel and incubated at room temperature for 1 ~ 2 h, or 4°C 2 ~ 4 h.
6. Centrifuge at 1000 rpm for 5 min and collect gel , remove unbound sample, and store for analysis.
7. Add 1000 μ L IP Lysis/Wash Buffer to the centrifuge tube and gently mix for 5 ~ 10 min. Collect the gel and discard the supernatant. Repeat twice.
8. **Denaturing elution:** Add 80 ~ 100 μ L SDS-PAGE Sample Loading Buffer (1 \times) to the centrifuge tube and heat the sample in 100°C water bath or metal bath for 10 min. Gel are separated by centrifugation to retain the supernatant containing the target antigen.

Note: The following elution methods can also be used to maintain protein activity.

Low pH Elution: Add 100 μ L Elution Buffer to centrifuge tube. Keep mixed and incubate centrifuge tube at room temperature for 5 ~ 10 min. The gel is separated by centrifugation to retain the supernatant containing the target antigen. Low pH is neutralized by adding 20 μ L Neutralization buffers per 100 μ L of wash out solution.



Notes

1. Before performing the experiment, please read this operation manual carefully.
2. The affinities between different types of antibodies and antigen Lysis in IP experiments are different. Lysis/Wash Buffer between antibodies and antigens is also affected by IP Lysis/Wash Buffer, so **do your own experiments by optimizing the operation details or screening and formulating buffers.**
3. Agarose gel should be fully oscillated and evenly before use. The gel should be kept in storage solution to prevent drying.
4. Transported at low temperature, Protein A/G agarose gel, IP Lysis/Wash Buffer, Elution Buffer and Neutralization Buffer are **stored at 4°C**. PMSF, Phosphatase inhibitor cocktail and SDS-PAGE Sample Loading Buffer were **stored at -20°C**.
5. This product is for scientific research only.

Problem Solving

1. No immunoprecipitation of antigen

- ① The sample contains too little antigen to be detected

Recommendation: To verify the expression and/or cleavage efficiency of proteins in the lysate by SDS-PAGE or protein immunoblotting; If necessary, increase the sample size

- ② Antibodies cannot bind to antigens

Recommendation: Choose another specific antibody, or choose another antibody that recognizes a different epitope.

- ③ Ingredients in IP Lysis/Wash Buffer interfere with the binding of antigens to antibodies

Recommendation: Immunoprecipitation and rinsing with other buffers (e.g., TBS containing 0.5% CHAPS)

2. Low protein intake

- ① The protein is degraded

Recommendation: Add protease inhibitor

- ② The amount of gel used is not enough

Recommendation: Increase the amount of gel used to capture immune complexes

- ③ The amount of target protein in the sample is not enough

Recommendation: Increase the amount of antigen sample

3. Multiple non-specific bands



There are non-specific proteins that bind to the gel

Recommendation: Add 50 ~ 350 mM NaCl to IP Lysis/Wash Buffer. Increase elution intensity and frequency.

4. Gel sticks easily to the tube wall

Recommendation: Use consumables with low adsorption rates for gel manipulation.

Ordering Information

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